The complete nucleotide sequence of RNA2 of barley mild mosaic virus (BaMMV)

ULRICH TIMPE and THOMAS KÜHNE

Federal Centre for Breeding Research on Cultivated Plants, Institute for Resistance Research, Theodor-Roemer-Weg 4, D-06449 Aschersleben, Germany

Accepted 20 April 1994

Key words: BaMMV, barley mild mosaic virus, bymovirus, nucleotide sequence, potyviridae, RNA2

Abstract. Barley mild mosaic virus is a member of the *Bymoviruses*, a genus of the family *Potyviridae*. The virus consists of two types of flexuous rod-shaped particles. Each of them contains one single-stranded polyadenylated RNA in plus orientation of approximately 7.6 kb (RNA1) and 3.6 kb (RNA2). Complementary DNAs of both RNAs have been synthesised and cloned. The nucleotide sequence of RNA2 has been determined. It is 3524 nucleotides in length, excluding the 3' poly(A) tail, and contains one large open reading frame (2679 nts), coding for a polyprotein of approximately 98 kDa. There are indications that a putative proteolytic activity in the N-terminal part can cleave the polyprotein autocatalytically into a 25 kDa protein (putative proteinase) and a 73 kDa polypeptide of unknown function.

Introduction

Barley mild mosaic virus (BaMMV) has been identified as one of the agents causing the yellow mosaic disease of winter barley. In nature this virus is transmitted by the soil-borne plasmodiophoid fungus *Polymyxa graminis* Ledingham [Adams et al., 1989]. The particles remain infective in resting spores of the fungus for many years. BaMMV possesses slightly flexuous rod-shaped particles with a bimodal length distribution of 270 to 289 nm and 568 to 600 nm [Huth et al., 1984]. Both genomic RNA molecules are single-stranded and have a poly(A) tail at their 3' terminus. At the 5' terminus they are expected to carry a genome-linked protein [Usugi et al., 1989]. RNA1 and RNA2 have molecular weights of approximately 2.5 MDa and 1.2 MDa, respectively. They are encapsidated by one type of capsid protein of about 35 kDa [Ehlers and Paul, 1986].

In winter barley BaMMV frequently occurs in mixed infections with barley yellow mosaic virus (BaYMV), which exhibits similar morphological and biological properties but can be differentiated serologically [Huth, 1986; Huth and Adams, 1990]. Recently, complete sequences of both RNAs of a Japanese and a German isolate of BaYMV have been published [Kashiwazaki et al., 1990, 1991; Davidson et al., 1991; Peerenboom et al., 1992], and the 3'-terminal part of BaMMV RNA1 has also been sequenced [Kashiwazaki et al., 1992; Schlichter et al., 1993].

Here we present the complete nucleotide sequence of RNA2 of a German isolate of BaMMV.

Materials and methods

Virus purification and RNA preparation

The isolate BaMMV-ASL1 originating from a field at Aschersleben (Germany) was propagated in a growth chamber on mechanically inoculated barley plants (cv. Erfa) and purified using the method of Huth et al. [1984]. Viral RNA was isolated from particles according to Kashiwazaki et al. [1989] with slight modifications (0.5% SDS, additional chloroform extraction step) and omitting the final oligo(dT) affinity chromatography.

cDNA synthesis and cloning

A mixture of both, RNA1 and RNA2 was transcribed into cDNA according to Gubler and Hoffman [1983] by using oligo(dT)primers. After synthesis of second strand DNA, treatment of the product with T4 DNA polymerase was made to generate blunt ends. Eco RI adapters were linked to the termini and the DNA fragments were ligated into bacteriophage λgt 11 arms and cloned in *E. coli* strain Y 1090 according to the instructions for the cDNA Cloning System Plus (Amersham, Braunschweig, Germany).

Two clones, L12 (1.2 kbp) and L27 (3.6 kbp) were identified by hybridisation from the genomic library which corresponded with BaMMV RNA2. After Eco RI digestion, the complete L12 insert and two L27 subfragments (0.8 and 2.8 kbp) were ligated into pGEM-3Zf(-) vector (Promega, Heidelberg, Germany), cloned and detected in both orientations as pG12(+) and pG12(-), pG2708(+) and pG2708(-), pG2728(+) and pG2728(-). To clone the complete insert of L27, a Bam HI fragment was ligated into the plasmid vector to give pG27. The extreme 5' terminus of RNA2 was cloned using the 5'-AmpliFINDER RACE kit (Clontech, Palo Alto, USA) and the virus specific cDNA primer 5'-GTGTGATATCAGCAGTGTCG-3' and the nested PCR primer 5'-ATGGAACTGCAGTGATCGGC-3' according to manufacturer's instructions. The 700 bp PCR product was cloned to give pG5'.

Sequencing

Both ssDNA and dsDNA sequencing was performed by the chain termination method [Sanger et al., 1977]. The clones, described above, but not pG27, and a set of restriction deleted subclones were used to determine overlapping sequences of both cDNA strands. In order to confirm the internal Eco RI single restriction site of pG27 a Hinc II-deleted subclone was also sequenced.

Results and discussion

The RNA2 of the isolate BaMMV-ASL1 comprises 3524 nucleotides (nts), excluding the 3' poly(A) tail (Fig. 1). The cloning of the extreme 5' terminus was performed with both a RNA preparation which had been stored for almost 3 years at -80 °C and a freshly prepared RNA. Following RACE PCR a number of independent clones, representing both RNA preparations was produced and sequenced. All preparations extended the 5' terminus of pG2708 by 14 nts and were identical in sequence. This confirmed that the extreme 5' terminus had been reached. However the possible presence of a covalently linked protein on the viral RNA might have hindered complete cDNA synthesis and precluded determination of the terminal nt.

Clone pG12 contains a track of about 80 A residues, indicating that the 3' terminus is covered by this clone.

The 140 nts long 5' untranslated region (UTR) is very rich in A and U, but poor in G similarly to other (+)RNA plant viruses, e.g., potyviruses. Conserved domains, which have been identified in the 5' UTR of potyviruses [Maiss et al., 1989; Turpen, 1989] are absent from both BaMMV RNA2 [this work] and BaYMV RNA2 [Davidson et al., 1991].

Recently, Kashiwazaki et al. [1991] reported about two short stretches of 7 nts (AAAGCAA and UUGCUUU) identical in the 5' region of BaYMV RNA1 and RNA2 that have the potential to base-pair. While some short sequences of potential base-pairing can also be identified by computer analysis in the 5' UTR of BaMMV RNA2, they do not resemble the sequences described for BaYMV.

BaMMV RNA2 contains a single large ORF between nts 141 and 2819. Four AUG start codons are present at the 5' terminus of the ORF, but only the first, starting at nt 141 is in good agreement with the consensus sequence for initiation of translation of eukaryotic mRNAs [Kozak, 1986; Staden, 1984]. The putative polyprotein encoded by this ORF comprises 893 amino acids and has a predicted molecular weight of 98 kDa. It shows only low similarity to the amino acid sequence of BaYMV RNA2. The most prominent homology is found in the N terminus of the polyprotein (Fig. 2). The amino acid motif GYCY, which occurs in the polyprotein encoded by the BaYMV RNA2, and in the HC-Pro of aphid-transmissible potyviruses, was identified to contain a proteolytically active cysteine [Oh and Carrington, 1989]. In BaMMV this domain has altered to GFCY and the putative active cysteine residue occurs at position 117. Furthermore, a histidine residue known as being essential for cleavage activity of HC-Pro of potyviruses has also been found in the polyprotein of BaMMV RNA2, at amino acid position 189. As in BaYMV, it is located 72 amino acids downstream of the cysteine residue of the GFCY motif. The surrounding amino acids of this histidine residue are nearly identical in the corresponding protein of the Streatley isolate of BaMMV [Andersen et al., 1993].

5 '	AAAAUAAAACUUCACAAAAAACCCAACGACUUAAAACGAACACAAUUAAGCAAACAAA	3 '
5 '	ACACCAAACGCACACCAACGAAUUACAGCACACAGCUGUAAAGCGCGAACUCAUUCACCGC 70 80 90 100 110 120	3'
5'	M M M N S M I R Q G W Q Q AAUCUUUGCAUUGAGCCAAGAUGAUGAUGAUCAUCAUGAUCCGCCAAGGAUGGCAGCAGG 130 140 150 160 170 180	13 3'
5 '	V L R R F S I P T S G D R L I V S N S T UGCUGAGGCGCUUUUCCAACUCCCAACAUCCGGAGAUAGACUCAUCGUUUCCAACUCCACCG 190 200 210 220 230 240	33 3'
5'	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	53 3'
5'	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	73 3'
5'	S V L E A S P R S F P W V F L T N S F C CCGUUCUUGAAGCAAGCCCACGGUCCUUCCCAUGGGUUUUUCUCACGAACUCAUUCUGCA 370 380 390 400 410 420	93 3'
5'	T F G G S I H A Q N L Q A F A T A E F K CCUUCGGUGGUUCGAUACAUGCUCAGAAUCUUCAAGCCUUCGCAACUGCCGAGUUUAAGA 430 440 450 460 470 480	113 3'
5,	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	133 3'
5'	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	153 3'
5'	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	173 3'
5 '	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	193 3'
5 '	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	213 3'
5'	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	233 3'
5'	Y S F W K K S M D H F T S D R F V E F L ACUCCUUCUGGAAGAAUCCAUGGACCACUUCACAUCGACCGGUUCGGAAUUCCUAG 850 890 900	253 3'
5 '	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	273 3'
5'	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
5'	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	
5'	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	

Fig. 1. Nucleotide sequence of BaMMV RNA2 and the amino acid sequence derived from the single large ORF starting at nucleotide 141.

I T H N V H Q H V F E V L K T M S V 5' CUGAUAUCACACACGUGCACCAGCACGUUUUUGAAGUCCUCAAGACCAUGUCUGUGC 3' F S K T T N A Y N R A R F E V N H K V 5' AGUUUAGCAAGACGAACGCCUACAACCGUGCCCGCUUUGAAGUCAACCACAAAGUCA 3' W N A E Y G R G P Q Q N A E L E A L V UAUGGAACGCAGAAUACGGACGCGGCCCUCAACAGAACGCCGAGCUCGAGGCUCUUGUAC 3 LFLNRQSLEIENILHRTTSP 5' UCUUUCUUAAUAGGCAAUCGCUUGAAAUUGAGAACAUCUUGCACAGAACAACUAGUCCUG 3' V V V T S W K P D V P P A A P E I K E E 43 UUGUUGUCACCAGCUGGAAACCCGAUGUUCCCCCUGCUGCACCUGAAAUCAAAGAGGAG 3 E P T H A I A T P I T E A P S H V T P V 5' AACCCACGCACGCAAUCGCAACACCAAUAACUGAGGCGCCAUCACAUGUCACUCCUGUUG 3' EVVNLPPTRSYWAETLVGIL AAGUUGUGAACCUGCCACCAACGCGCUCCUAUUGGGCUGAGACACUUGUCGGUAUCCUCA 3 I L G T V F A F L T R A L I R A K R CUGCAAUCCUGGGAACGGUAUUCGCCUUCCUCACUCGAGCACUCAUUCGUGCAAAGAGAU 3' L R R K S T F P W V T L N S G D D D D 5' UGCGGAGGAAAUCCACCUUCCCCUGGGUGACGCUAAACUCUGGAGAUGACGAUGAUGACC 3' Q S G G G G G P Q T P G G Q P P V P H 53 1710 1720 1730 TRGTHQSRFSVODIASDTSL 5' CGCGUGGAACGCACCAGUCGCGCUUCUCCGUGCAGACAUAGCAUCCGACACCAGUCUGC 3' LSVDLDEDTLSQYDETFQKI UAAGUGUCGAUCUCGACGAGGACACGCUCUCCCAGUAUGAUGAAACCUUCCAAAAGAUCC 3' R R A L F E T S F A D I L Q N S A R W I 59 GCCGUGCGUCUUUGGAAACCAGUUUUGCAGACAUCCUGCAAAACUCUGCUCGUUGGAUCU 3 TLEAMALADGNAPYTLLAQ CCACGCUGGAAGCGAUGGCUCUCGCAGAUGGCAAUGCUCCUUACACACUUCUUGCGCAGU 3 Y L N G I E E A Y T N F R N T G H I S R AUCUCAACGGGAUUGAGGAGGCAUACACGAACUUUCGCAACACAGGUCACAUCUCCCGCG 3' ATLSGFFALEDNLR A AGT 5' CAACACUUUCAGGCUUCUUCGCCUUGGAAGAUAAUCUGCGUGCAGCUGGCAUAGCUUUUG 3' PTQTIQNQFADSPARR 5' GGACCACAACACCCACGCAAACCAUACAGAACCAAUUUGCCGACUCCCCAGCCCGUCGAU 3 KTRFEQIACELGDASIKSL 5' GGAAAACACGAUUUGAACAGAUUGCAUGUGAACUGGGCGAUGCUAGCAUCAAAUCGCUUG 3' D L A D I I D T E R E R G D L T Q F D CAGAUCUUGCAGACAUCAUCGACACUGAGCGGGAGAGGGGCGACUUGACUCAGUUUGAUG 3

Fig. 1. Continued.

5 '	OUCUC V L	A GCA 22		s s		I UC#			L CUCU 2310		R CGU(A SCU(23)	R CGC#		I AUC 330		D GAG	T CACCA 2340	733 3'
5'	T D CCGAU	P CCA 23		A GCU		L IUAC 160		UU	V 3000 2370		N AAC	A SCAI 238	A GCU <i>I</i>		Q CAG 390	N AAC	N AA	I CAUCA 2400	753 3'
5'	N A ACGCC	I AUA 24		G GGC1		N ACC 20			I AUAC 2430		F JUCO	L CUCI 24	A GCG <i>I</i>		R CGU 450	R CGU	L UUI	L ACUCA 2460	773 3'
5 '	I T UCACA	R CGC 24		y Gបបថ		E AAC 80			A GCGC 2490		S AGCC	R CGCU 250	G GGAG		T ACG 510	CCU	E GA/	T AACCG 2520	793 3'
5′	V Q UUCAG	Q CAA 25		A GCU		A CAC			A 3CA# 2550		I AUUC	V SUG! 256	E		N AAC 570	M AUG	Y UA(N CAAUG 2580	813 3'
5'	E M AGAUG	A GCC 25		S AGC		R GGC 00			A GCC# 2610		A GCA#	T ACAG 262	E GAAC		T ACC 630	I AUC	R CGC	E CGAGC 2640	833 3'
5 '	H V ACGUG	L CUC 26		CCC		N ACG 60			A GCA# 2670		v รบบด	G GC/ 268	A GCUG	CU	A GCG 690	F UUC	F UU(R CGAU 2700	853 3'
5 '	s G CCGGU	G GGU 27		R CGCt		R GGG			N AACC 2730		A GCAI	M SUGO 274	T ACAP		P CCC 750	G GGU	G GGI	P JCCUG 2760	873 3'
5'	A A CUGCU	A GCG 27		R CGG(M UGU			A 3000 2790		R AGGC	G GA0 280	G GGGC		R CGU 810	CUU	n AAU	R JAGGU 2820	893 3'
5'	* AGGCA	UCA 28		cuci		CUC 40	UGG		CAGC 2850		JGAI	ACG? 286	GACA		CGC 870	UCG	טטכ	2880	3,
5'	GAUCC	AAU 28		ACG		GCG	UGC		CUGA 2910		AAA	GAF 292	ACGU		GUU 930	GUG	AUA	AUUA 2940	3'
5'	AAUUA	ນບບ 29	-	CGCI		เบบบ 60	GUC		JGUU 2970		CUAC	298	CUC		CUG 990	CAG	CUC	3000	3'
5'	GGAGG	30 30		CGG		GAC 20	AUC		CACA 3030		CCAt	304	AACC		CUG 050	CAU	UCI	ACAUC 3060	3'
5 '	บบบเรบ	GCC 30		UUAI		CUC 180	AUA		309C		CAAC	310	JUCU		CAG 110	AGC	AUC	3120	3'
5'	AGCAU	CGA 31		UCAG		AUU 40	IGCA.		3150		CUU	316	CCGG		սնց 170	CAA	UCF	3180	3'
5'	UGAGU	ՍԸՍ 31		ucco		GUG OO	UUC		20GC		CUGO	322	JCGU		บูตูบ 230	UGU	CGC	UGAU 3240	3'
5'	UACUAU	JCAG 325		GUA	320		GAUA		CAC(270	CCA		UCA 328	ACU		GUA 90	CUZ		agua 3300	3'
5′	CCGGAC	331		JAUC	33:		UUAA		CUG 330			GCU 334	AGG		GC0 150	AUS		UUAC 3360	3'
5'	CAUUUG	337		ΙΌŪG	338		cucc		CUC 390			AAA 340	AGU:		UGC 10	AUA		AAAC 3420	3 '
5'	GUCAUC	343		CUGU	UCA 34		UGAG		CGG ¹ 450			AAG 346	CAA	GUC 34	AUC 170	CAU	AGA	CUGC 3480	3 '
5'	UGGCAU	JUGU 349		GUU	GAC 35		GUUU		GUA 510			AAC 352	AC .	3 '					

Fig. 1. Continued.

					-				71												
Bammv	112	F	K	s		F	C *	Y	м	N	L	L	I *	P	L	s	F	D	I	I T	D
BayMV	138	*	A	Н	<u> </u>	Y	*	*	ļr	s	*	F	*	*	*	*	*	R	*	T	P
		A	Н	A	Đ	s	F	R	G	F	V	E	Q	L	P *	D	T	Ļ	G		Y
		E	N	*	R	*	*	S	R	*	L	*	*	*	*	*	Ι	*	*	*	*
		P	s		s	M	v	L	N	v	М	L	H	A	A	T	R L	F *	P *	E	I V
		*	Т	*	A	s	L	Y	K	T	*	*	F	*	v	R	L	*	*	*	V
		v	A	s	P	I	P	T	I	Α	F	D	A	E	s V	L	Q	F	н	v *	T
		L	Q	A	P *	*	*	Ι	*	*	K	R	P	G	V	*	*	*	<u> </u>	*	s
		D	K	R	G	V	P	G	М	W	N	1	L	ĸ	A	С	R	V	Y	E	L
		*	A	*	*	L	*	P	s	*	F	P	М	*	C	G	s	*	A	s	F
		L	s	L	A	A	D	G	I	G	C	E	Y	М	L	Y	P	v	G	A	230
		1	A	*	1	T	N	N	L	N	S	D	L	L.	N	G	P	*	*	s	256
																		_			

Fig. 2. Comparison of amino acids 112 to 230 of BaMMV RNA2 with the corresponding region of the polyprotein of the German isolate of BaYMV [Davidson et al., 1991]; amino acid residues identical for BaMMV and BaYMV are marked by asterisks (*) in the BaYMV sequence; boxed sequences represent the proteolytically active regions of the putative proteinase and the putative cleavage site of the polyprotein.

The putative G/S cleavage site in the polyprotein of BaYMV RNA2 is replaced by a G/A dipeptide in BaMMV at positions 229/230. In addition, as for potyviruses and BaYMV the G is preceded by a valine.

Cleavage at this site will lead to a 25 kDa protein, a putative proteinase (28 kDa in BaYMV) and a 73 kDa protein of unknown function (70 kDa in BaYMV). No other ORF, starting with an AUG and a coding capacity for a peptide larger than 12 kDa, could be detected.

The 3' non-coding region of BaMMV RNA2 is 705 nts long and, as shown in Fig. 3, there is a good homology between the German isolate and the published 141 nts from the extreme 3' terminus of RNA2 of the Streatley isolate of BaMMV [Andersen et al., 1993]. The 3' non-coding region contains the sequence UAUGU at positions 3505 to 3509. This sequence was originally described at the DNA level to be a polyadenylation signal in yeast and necessary for transcription termination [Zaret and Sherman, 1982]. It was first detected in the sequence of a plant virus RNA by Maiss et al. [1989] and similar putative signals have been identified in a number of other plant virus RNAs. However, as yet the proposed function

Fig. 3. Comparison of the nucleotide sequence of the 3'-terminal parts of RNA2 of the BaMMV isolates from (a) Aschersleben, Germany and (b) Streatley, U.K. [Andersen et al., 1993].

of this signal has not been confirmed experimentally. Infectious full-length clones should be a good tool to elucidate the biological function of this sequence.

The nucleotide sequence data reported here will appear in the EMBL, GenBank and DDBJ Nucleotide Sequence Databases under the accession number X75933.

Acknowledgements

We should like to thank Mrs Doris Walther for excellent technical support and Drs H. Huttinga, F. van der Wilk and G. Proeseler for their engagement at the initial stage of this work. UT was granted by the Deutsche Forschungsgemeinschaft (Pr 420/1-1).

References

- Adams MJ, Batista MF, Swaby AG and Antoniw JF (1989) Fungally-transmitted viruses of cereals in the UK. Eppo Bull. 19: 573-577
- Andersen JF, Davies JW and Coutts RHA (1993) Further evidence for the inclusion of barley mild mosaic virus (BaMMV) in the baymovirus group. J. Phytopathology 139: 48-56
- Davidson AD, Pröls M, Schell J and Steinbiss H-H (1991) The nucleotide sequence of RNA2 of barley yellow mosaic virus. J. gen. Virol. 72: 989-993
- Ehlers U and Paul H-L (1986) Characterization of the coat proteins of different types of Barley yellow mosaic virus by polyacrylamide gel electrophoresis and electro-blot immunoassay. J. Phytopathology 115: 294-304
- Gubler U and Hoffman BJ (1983) A simple and efficient method for generating cDNA libraries, Gene 25: 263-269
- Huth W (1986) Isolierung mehrerer Stämme des Gelbmosaikvirus der Gerste (barley yellow mosaic virus, BaYMV). Mitt. Biol. Bundesanstalt Berlin 232: 384
- Huth W and Adams MJ (1990) Barley yellow mosaic virus (BaYMV) and BaYMV-M, two different viruses. Intervirology 31: 38-42
- Huth W, Lesemann D-E and Paul H-L (1984) Barley yellow mosaic virus: purification, electron microscopy, serology and other properties of two types of the virus. Phytopath. Z. 111: 37-54
- Kashiwazaki S, Hayano Y, Minobe Y, Omura T, Hibino H and Tsuchizaki T (1989) Nucleotide sequence of the capsid protein gene of barley yellow mosaic virus. J. gen. Virol. 70: 3015-3023
- Kashiwazaki S, Minobe Y and Hibino H (1991) Nucleotide sequence of barley yellow mosaic virus RNA2. J. gen. Virol. 72: 995-999
- Kashiwazaki S, Minobe Y, Omura T and Hibino H (1990) Nucleotide sequence of barley yellow mosaic virus RNA1: a close evolutionary relationship with potyviruses. J. gen. Virol. 71: 2781-2790
- Kashiwazaki S, Nomura K, Kuroda I and Hibino H (1992) Sequence analysis of the 3'-terminal halves of RNA1 of two strains of barley mild mosaic virus. J. gen. Virol. 73: 2173-2181
- Kozak M (1986) Point mutations define a sequence flanking the AUG initiator codon of eucaryotic ribosomes. Cell 44: 283-292
- Maiss E, Timpe U, Brisske A, Jelkmann W, Casper R, Himmler G, Mattanovich D and

- Katinger HWD (1989) The complete nucleotide sequence of plum pox virus RNA. J. gen Virol. 70: 513-524
- Oh C-S and Carrington JC (1989) Identification of essential residues in potyvirus proteinase HC-Pro by site-directed mutagenesis. Virology 173: 692-699
- Peerenboom E, Pröls M, Schell J, Steinbiß H-H and Davidson AD (1992) The complete nucleotide sequence of RNA1 of a German isolate of barley yellow mosaic virus and its comparison with a Japanese isolate. J. gen. Virol. 73: 1303-1308
- Sanger F, Nicklen S and Coulson AR (1977) DNA sequencing with chain-terminating inhibitors. Proc Natl. Acad. Sci. USA 74: 5463-5467
- Schlichter U, Sohn A, Peerenboom E, Schell J and Steinbiß H-H (1993) Molecular analysis of the capsid protein gene of a german isolate of barley mild mosaic virus. Plant Cell Rep. 12: 237-240
- Staden R (1984) Computer methods to locate signals in nucleic acid sequences. Nucl. Acids Res. 12: 505-519
- Turpen T (1989) Molecular cloning of a potato virus Y genome: nucleotide sequence homology in non-coding regions of potyviruses. J. gen Virol. 70: 1951–1960
- Usugi T, Kashiwazaki S, Omura T and Tsuchizaki T (1989) Some properties of nucleic acids and coat proteins of soil-borne filamentous viruses. Ann. Phytopathol. Soc. Jpn. 55: 26-31
- Zaret KS and Sherman F (1982) DNA sequence required for efficient transcription termination in yeast. Cell 28: 563-573